

JOURNAL OF THE WORLD RABBIT SCIENCE ASSOCIATION

# July - September 2021 World Rabbit Sci. 29 (3) 129 - 201



World Rabbit Science Association



UNIVERSITAT Politècnica de València



JOURNAL OF THE WORLD RABBIT SCIENCE ASSOCIATION

# July - September 2021 World Rabbit Sci. 29 (3) 129 - 201



World Rabbit Science Association



UNIVERSITAT POLITÈCNICA DE VALÈNCIA



JOURNAL OF THE WORLD RABBIT SCIENCE ASSOCIATION

#### Editor in Chief

#### J.J. Pascual

Universitat Politècnica de València, Instituto de Ciencia y Tecnología Animal. P.O. Box 22012, 46071 Valencia. Spain. jupascu@dca.upv.es

#### Associate Editors

#### J.M. Corpa

Instituto CEU de Ciencias Biomédicas, Departamento PASAPTA. Facultad de Veterinaria. Universidad Ceu Cardenal Herrera C/ Tirant lo Blanch, 7. 46115 Alfara del Patriarca, Valencia. Spain. imcorpa@uchceu.es

#### P. García

Dpto. Producción Agraria.- Campos de Prácticas. Escuela de Ingeniería Agronómica, Alimentaria y de Biosistemas Universidad Politécnica de Madrid. Cdad. Universitaria, s/n. 28040 Madrid. Spain. pilar.grebollar@upm.es

#### H. Garreau

UMR 1388 GenPhySE : Génétique, Physiologie et Systèmes d'Elevage 24 Chemin de Borde Rouge. Auzeville Tolosane, CS 52627. 31326 Castanet Tolosan Cedex. France. herve.garreau@toulouse.inra.fr

#### G. González-Mariscal

Centro de Investigación en Reproducción Animal, CINVESTAV-UAT. Apdo Postal 62, Tlaxcala, Tlax. 90000, Mexico. gqlezm@prodigy.net.mx

#### P. Hernández

Universitat Politècnica de València. Instituto de Ciencia y Tecnología Animal. Camino de Vera s/n. 46071 Valencia. Spain. phernan@dca.upv.es

#### M. Petracci

Dipartimento di Scienze e Tecnologie Agro-Alimentari Alma Mater Studiorum Università di Bologna Piazza Goidanich, 60. 47521 Cesena. Italy. m.petracci@unibo.it

#### Z. Volek

Department of Nutrition Physiology and Animal Product Quality. Institute of Animal Science. Přátelství 815. CZ-104 00, Prague-Uhříněves. Czech Republic. volek.zdenek@vuzv.cz

#### ASSOCIATE DIRECTOR

#### S. Calvet

Universitat Politècnica de València. Instituto de Ciencia y Tecnología Animal. Camino de Vera s/n. 46071 Valencia. Spain. salcalsa@upvnet.upv.es World Rabbit Science is the official journal of the World Rabbit Science Association (WRSA). One of the main objectives of the WRSA is to encourage communication and collaboration among individuals and organisations associated with rabbit production and rabbit science in general.

World Rabbit Science is the only international peer-reviewed journal included in the ISI Thomson Scientific devoted to publishing original research in the field of rabbit science. World Rabbit Science is indexed and abstracted in the SciSearch®, Current Contents® and Focus On (Veterinary Science & Medicine) since the beginning of 2005. Papers or reviews of the literature submitted to World Rabbit Science must not have been published previously in an international refereed scientific journal. Previous presentations at a scientific meeting, field day reports or similar documents can be published in World Rabbit Science, but they will be also subjected to peer-review process.

World Rabbit Science is published in English four times a year in a single volume. Authors may publish in World Rabbit Science regardless of the membership in the World Rabbit Science Association, although joining the WRSA is highly encouraged. Views expressed in papers published in World Rabbit Science represent the opinion of the author(s) and do not necessarily reflect the official policy of the WRSA or the Editor in Chief.

#### EDITORIAL OFFICE

World Rabbit Science Instituto de Ciencia y Tecnología Animal Universitat Politècnica de València. Camino de Vera s/n. 46071 Valencia. Spain

#### EDITORIAL SECRETARY

C. Lario, Managing Editor Contact for questions on relations with Section Editors, submissions and proofs. Universitat Politècnica de València. Editorial UPV Camino de Vera s/n. 46071 Valencia. Spain. colarma@editorial.upv.es

#### SUBSCRIPTION INFORMATION

Volume 29, 2021, 4 issues: 80  $\in$  For more information, contact the Editorial UPV: pedidos@editorial.upv.es

#### Published by

Universitat Politècnica de València. Camino de Vera s/n. 46071 Valencia. Spain. Instructions to authors are available at:

#### www.wrs.upv.es

This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License



#### World Rabbit Science 2<sup>nd</sup> Series

PRINTED IN SPAIN BY: Byprint percom, s.l. LEGAL DEPOSIT: nº V-1162-2003, Marzo 2003. COVER DESIGN: A. Climent. Universitat Politècnica de València. LAVOUT: Enrique Mateo. Triskelion Disseny Editorial. ISSN: 1257-5011 EISSN: 1989-8886









# HIERARCHICAL CLUSTERING AS A TOOL TO DEVELOP A CLASSIFICATION SCHEME FOR RABBIT MEAT QUALITY

LARIVIÈRE-LAJOIE A.-S.\*\*, CINQ-MARS D.<sup>10</sup>\*, GUAY F.<sup>10</sup>\*, BINGGELI S.<sup>10</sup>\*, DALMAU A.<sup>10</sup>\*, SAUCIER L.<sup>10</sup>\*\*

\*Department of Animal Science, Faculty of Agriculture and Food Science, Université Laval, QUEBEC CITY, Quebec, Canada, G1V 0A6. <sup>†</sup>Institute of Nutrition and Functional Foods, Université Laval, QUEBEC CITY, Quebec, Canada, G1V 0A6. <sup>‡</sup>Institute of Agrifood Research and Technology (IRTA), MONELLS, 17121, Girona, Spain.

Abstract: This study aimed to characterise the quality of meat from commercially-raised rabbits. Animals came from five different producers and were slaughtered in three different plants under provincial or federal inspection jurisdiction. Animal behaviour evaluated by scan sampling prior to feed withdrawal (FW) and transport, as well as blood lactate concentration at exsanguination, did not raise concerns with respect to stress. Stomach pH was higher (P=0.047) when the FW time was short ( $\leq 13.5$  h), at a mean value of 2.23. All pH values measured 1 h post-mortem from the Biceps femoris (BF) and almost all (97.6%) from the Longissimus lumborum (LL) were higher than 6. Values for ultimate pH measured 24 h postmortem (pH,) ranged from 5.80 to 6.83 and from 5.70 to 6.70 for BF and LL muscles, respectively. The maximum meat drip loss recorded was 2.6%, while cooking loss reached 30%. Meat lightness (L\*) and colour intensity (C\*) for the long FW times (≥23 h) were no different from those with short and intermediate (15.5 to 17.3 h) FW times. However, these colour parameters were higher for the short FW time class compared to the intermediate FW time class (P<0.02). A hierarchical cluster analysis based on pH, cooking loss and lightness (L\*) from 200 rabbit loins was performed. Of the four clusters created, clusters 1 and 2 had the best and second-best meat quality, respectively. Clusters 3 and 4 had the lowest meat quality and presented DFD-like (dark, firm and dry) characteristics. Meat did not exhibit PSE-like (pale, soft, exudative) characteristics, even for the slaughter lot with the minimum mean pH... Of the eight slaughter lots evaluated, more than 50% of the meat from three of them fell into clusters 3 and 4; all three were in the intermediate FW time class. Overall, the quality of rabbit meat analysed was acceptable for commercial use, but rather variable. This suggests that there are factors within the value chain that are not yet fully controlled and require further investigation.

Key Words: cooking loss, DFD meat, feed withdrawal, hierarchical cluster analysis, pre-slaughter management, rabbit.

# INTRODUCTION

According to the FAO (2017), global production of rabbit meat worldwide was about 1.4 million tonnes and came principally from Asia (75.3%), followed by Europe (21.3%), Africa (7.1%) and America (1.2%). Of the 1.2% produced in America, Canada and the US produced 1.03%. In 2016, the provinces of Ontario and Quebec were responsible for 55.32% and 35.49% of Canadian rabbit meat production, as 33.47% and 18.99% of the rabbit farms are located in these provinces, respectively (AAFC, 2019). Since 2011, rabbit production has been relatively stable in Quebec, partly because it is considered to be a specialty meat and is often associated with holiday celebrations. In 2014, consumption was relatively low at 0.0254 kg per capita in Canada (AAFC, 2019) and 0.040 kg in the Quebec province (MAPAQ, 2015). However, from 2010 to 2014, rabbit meat consumption increased to 3% per year in Quebec alone (MAPAQ, 2015). Although the agri-food activity of rabbit production is marginal in Quebec and in Canada in general, it

Correspondence: L. Saucier, *linda.saucier@fsaa.ulaval.ca.* Received *September 2020* - Accepted *April 2021*. https://doi.org/10.4995/wrs.2021.14368



contributes to the diversity of the food supply and its development can only be achieved if farmers produce adequatequality meat.

Meat quality defects can cause economic losses for both producers and processors, and may cause a reduction in meat shelf life (Faucitano *et al.*, 2010; Adzitez and Nurul, 2011). Animals that are exposed to acute or chronic stress just before slaughter can yield either pale, soft and exudative (PSE) or dark, firm and dry (DFD) meat, respectively, which are two well-known meat quality defects (Adzitey and Nurul, 2011). The incidence of PSE meat reduces processing yield, and the high microbial spoilage associated with DFD meat decreases its shelf life (Faucitano *et al.*, 2010; Saucier, 2016; Ponnampalam *et al.*, 2017). Furthermore, because two of the most important variables driving consumer choice are the colour and consistency of raw meat (Dalle Zotte, 2002), PSE and DFD raw meats have lower value compared to normal meats due to their unattractive appearance (Viljoen *et al.*, 2002).

Meat quality is influenced by many factors including pre-slaughter management, such as feed withdrawal (FW), transport and lairage time. Pre-slaughter FW is important for reducing transport-related sickness, incidence of downers and death, as well as microbial contamination during transport (Martin-Peláez *et al.*, 2008). Furthermore, FW reduces the volume of the gastrointestinal tract, in particular the stomach, which also reduces puncture risk during evisceration (Dalle Zotte, 2002). However, previous studies have established that extended FW could result in a reduction in live body weight (Bianchi *et al.*, 2008; Frobose *et al.*, 2014). Within the first hours of FW, stomach weight reduction is observed, while a prolonged FW can cause degradation of body tissues, loss of nutrients and humidity, leading in turn to quality and yield losses (Bianchi *et al.*, 2008). An extended FW period is ascribed to poor animal welfare conditions as expressed by an increase in aggressivity, with longer and more intense fighting (Faucitano *et al.*, 2006), and can reduce the levels of muscular glycogen reserves, which can lead to undesirable meat with a high pH (Faucitano *et al.*, 2006; Verga *et al.*, 2009). Furthermore, it has been reported that longer transport journeys increase bruising and mortality (Petracci *et al.*, 2008).

Pork is classified into five quality categories based on the ultimate pH (pH<sub>u</sub>), colour and meat drip loss: PSE, PFN (pale, firm, non-exudative), RSE (reddish-pink, firm, exudative), RFN (reddish-pink, firm, non-exudative; normal pork) and DFD (Faucitano *et al.*, 2010). For beef, the classification method considers carcass yield and texture, but also colour in order to identify DFD meat (Polkinghorne and Thompson, 2010; Ponnampalam *et al.*, 2017). Traditionally, PSE meat was associated with pigs and DFD meat with all species (Adzitey and Nurul, 2011). However, PSE-like meat has now been identified in turkey, chicken and cattle (Adzitey and Nurul, 2011). DFD-like meats have been reported in the literature for rabbits (Jolley, 1990; Koné *et al.*, 2016; Składanowska-Baryza *et al.*, 2018), whereas PSE-like meat has not (Cavani *et al.*, 2009; Blasco *et al.*, 2018).

To our knowledge, no formal and specific meat quality classification has ever been defined for rabbit meat. Despite the implementation of a code of good practices (RMAAQ, 2019), meat quality may vary. Thus, the aim of this study was to characterise meat quality from rabbits commercially raised in Quebec and slaughtered in facilities under federal or provincial jurisdiction. Using a hierarchical cluster analysis based on  $pH_u$ , cooking loss and lightness (L\*), a classification of rabbit meat quality is proposed.

# MATERIALS AND METHODS

All experimental procedures involving live rabbits were approved by Université Laval's Animal Use and Care Committee, which strictly adheres to the Guidelines of the Canadian Council on Animal Care (CCAC, 2009). Rabbits were commercially raised and analyses were performed from January 2018 to August 2019.

# Producer selection

Five rabbit producers were selected in collaboration with the *Syndicat des producteurs de lapins du Québec* to represent the vast majority of procedures in operation within the province of Quebec. Rabbits were slaughtered in three different abattoirs located in either Ontario or Quebec. They operated under provincial or federal inspection. The pre-slaughter management practices are presented in Table 1 for each slaughter lot.

								Total feed
Slaughter lot		Inspection		Rabbits	Feed withdrawal	Transport	Lairage	withdrawal
designation	Producers	juridiction <sup>2</sup>	Season	per lot	time at the farm (h)	time (h)	time (h)	time (h) <sup>3</sup>
A-P1-W	А	Provincial 1	Winter	270	6.66	0.17	1.67	8.5
B-F1-W	В	Federal 1	Winter	450	2.5	5	19	26.5
C-F1-W	С	Federal 1	Winter	900	10	5	14	29.0
D-P1-W	D	Provincial 1	Winter	747	9.67	2.25	3.58	15.5
D-P1-S	D	Provincial 1	Summer	800	9.75	2	1.75	13.5
D-P2-S	D	Provincial 2	Summer	805	12.75	2.25	1	16.0
D-F2-S	D	Federal 2	Summer	760	12.75	2.25	2.33	17.3
E-F1-S	E	Federal 1	Summer	320	0	7	16	23.0

Table 1: Pre-slaughter management according to the producer and the season<sup>1.</sup>

<sup>1</sup>Pre-slaughter management varies according to which abattoir the rabbits were delivered.

<sup>2</sup>Indicates the slaughterhouse inspection jurisdiction under which rabbits were slaughtered. Provincial 1 and 2 were located in Quebec; Federal 1 was located in Ontario; Federal 2 is the same slaughterhouse as Provincial 2, but after it received federal accreditation.

<sup>3</sup>Total feed withdrawal time includes all time segments: feed withdrawal time while at the farm, during transport and in lairage at the abattoir.

# Behavioural observations

For each slaughter lot (n=8), animal behaviour was evaluated by visual scan sampling at one minute intervals for a total of 10 min using an observation grid on 10% of the total cages prior to FW. The number of rabbits sitting, lying down or moving was recorded. Observations also included occurrence and types of activities and interactions. Aggressive behaviours relate to chasing and triggering escape, leaping, biting another rabbit, bouncing and paw scraping. All behavioural assessments were performed by the same observer.

## Physiological measures

For each lot slaughtered in the abattoirs located in Quebec, 25 rabbits were randomly selected from the dressing line. Blood samples were collected at exsanguination to measure blood lactate level, in duplicate, using a hand-held lactate analyser (Lactate scout +, EKF Diagnostics, Cardiff, Wales, UK) according to the manufacturer's specifications.

Full gastrointestinal tracts (GIT) were promptly removed and weighed after slaughter. Stomachs and caeca were tied at both ends, removed and weighed when full and then again when empty. The pH of the caecum and stomach contents was measured using a portable pH meter (ROSS, Orion Star A221, Thermo Scientific, Beverly, CA, USA) combined with an Orion Kniphe electrode (ThermoFisher, Nepean, ON, Canada) and a temperature compensation probe (928,007 MD, micro probes ATC, Maryland, USA). Stomach and caecum contents were weighed and kept at –20°C until the percentage of dry matter (DM) was evaluated. DM was calculated after lyophilisation (model 50L Virtual EL-85, VirTis, Los Angeles, CA, USA) at 20°C for 3 d. The water content was determined by calculating the weight difference between the wet and dry contents (Saucier *et al.*, 2007).

# Meat quality measurement

To determine the meat quality, the rabbit carcasses were analysed according to Koné *et al.* (2019). Muscular pH of the *Longissimus lumborum* (LL) and the *Biceps femoris* (BF) muscles were measured 1 h (pH1h) *post-mortem* for animals slaughtered in the abattoirs located in Quebec (since we had access to the processing line), and after 24 h (ultimate pH (pH<sub>u</sub>)) with a portable pH meter (ROSS, Orion 4 Star, Thermo Scientific) combined with an Orion Kniphe electrode (ThermoFisher) and an Orion<sup>TM</sup> Stainless-Steel Automatic Temperature Compensation (ATC) Probes (#927007MD, Thermo Scientific; Blasco and Ouhayoun, 1996). Meat colour was measured 24 h after slaughter on LL muscle cross sections between the 6<sup>th</sup> and 7<sup>th</sup> lumbar vertebrae (Dalle Zotte *et al.*, 2015) and on the exposed surface overlying the BF (Dalle Zotte *et al.*, 2009). After exposing the cut muscle surface to ambient air for 20 min ("blooming time"; Koné *et al.*, 2019), meat colour was evaluated using a Chroma meter (CR 400, Minolta Ltd., Osaka, Japan) equipped with a conical open port and an 8 mm aperture, a diffuse illumination/0° viewing angle geometry and a D65 light source

according to the reflectance coordinates (L\*, a\*, b\*; CIE, 1976). Parameters used to compare meat colour were lightness (L\*), redness (a\*), yellowness (b\*), colour intensity (chroma, C\*) and the hue angle (h). Equation (1) was used to calculate the chroma, while Equation (2) was used to determine the hue angle (Pathare *et al.*, 2013):

$$C^{*} = \sqrt{(a^{*2} + b^{*2})}$$
(1)  
$$h = Tan^{-1}(\frac{b^{*}}{a^{*}})$$
(2)

For samples with a negative a\* value, 180° was added to the calculated h value (McLellan *et al.*, 1995). Drip loss was measured by the weight difference of a piece of LL (2 cm thick×2.5 cm in diameter) after storage at 4°C for 48 h using an EZ-Driploss cup (Meat Extract Collector, Sarstedt AG & Co. KG, Nümbrecht, Germany; Rasmussen and Anderson, 1996). The cooking loss was evaluated using a similar piece of LL muscle (Pla, 1999) and is expressed as a percentage of the initial weight loss. Samples were placed individually into a Whirl-Pak bag (S-19793, Nasco Whirl-Pak®, USA), the air was removed from the bag, and it was then submerged in a water bath at 70°C for 15 min. Samples were then cooled in an ice-water bath, removed from the bag and weighed after removing the excess moisture with filter paper (Vergara *et al.*, 2005).

# Statistical analysis

To determine the differences in the behavioural parameters between slaughter lots, data were assessed using the SAS (Statistical Analysis System, SAS Institute Inc. 2002) GLIMMIX procedure. The LSMEANS statement adjusted by a Tukey's test was used to compare the differences between slaughter lots. In a second analysis, season was used as a fixed effect in order to evaluate its impact; producer was used as a random variable.

For the physiological and meat quality parameters, data were analysed using the SAS GLIMMIX procedure. To measure the effect of FW time, three classes were established according to what was applied by the selected producers (class 1, short $\leq$ 13.5 h; class 2, 13.5 h<intermediate<23 h; and class 3, long $\geq$ 23 h; Table 1). FW time, class and season were used as fixed variables, whereas slaughter lot and slaughterhouse were random variables.

Pearson's correlation coefficients were calculated on the residuals of lot analysis to measure the associations between the parameters under study using JMP 15 (SAS Institute Inc. 2002). For the principal component analysis (PCA), only the lots that were slaughtered in the abattoirs located in Quebec were considered. This was because access to the processing line was denied for lots slaughtered in the Ontario abattoir, therefore preventing the collection of physiological data. SAS software was used for this analysis.

A hierarchical cluster analysis was performed using Minitab software (Release 19) to identify groups of rabbit meat with different quality characteristics. Between each pair of observations, the Euclidian distance was used to measure the resemblance between groups and a complete linkage clustering method was used to associate similar samples. Four clusters were formed based on three meat quality variables ( $pH_u$ , cooking loss and L\*). An analysis of variance (ANOVA) using the SAS software MIXED procedure was performed to evaluate differences between quality characteristics of the groups formed by the cluster analysis, and the LSMEANS statement adjusted by a Tukey's test was used to compare the differences between clusters.

# RESULTS

# Behavioural parameters

According to Figure 1, only a small percentage of rabbits were observed to be moving ( $\leq$ 3.1%), whereas most were sitting or lying down. Rabbits from A-P1-W were documented as moving more frequently than those from E-F1-S. Rabbits from D-P2-S, A-P1-W and D-P1-W were found to be sitting more often (60.6, 53.5 and 51.9%, respectively) than rabbits from D-P1-S and E-F1-S (35.7 and 32.7%, respectively). Values obtained for the other slaughter lots ranged from 39.8 to 51.5%. Results for rabbits lying down were essentially the opposite of those sitting.

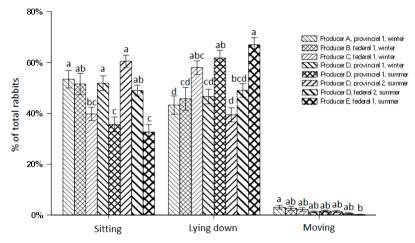


Figure 1: Percentage of rabbits sitting, lying down or moving during observations, prior to feed withdrawal, at each farm visit (mean $\pm$ standard error). Bars with different letters differ significantly at P<0.05.

For all slaughter lots, limited interaction between rabbits prior to FW was observed (Table 2). However, the proportion of rabbits exhibiting nonaggressive interactions was higher (P=0.046) for C-F1-W (4.0%) compared to D-P1-S (1.0%; Table 2). A-P1-W was the only slaughter lot where rabbits expressed aggressive behaviour (biting another rabbit; 0.4%).

D-P1-W had more rabbits resting than A-P1-W (89.0 vs. 79.8%; P=0.06; Table 2). Slaughter lot D-P2-S (3.8%) had more rabbits that were drinking than B-F1-W (0.8%), C-F1-W (1.2%) and E-F2-S (0.9%; all *P*-values<0.005; Table 2). A-P1-W, E-F2-S, D-P1-S, D-P2-S and D-F2-S had more rabbits that were grooming than D-P1-W (all *P*-values<0.049; Table 2).

B-F1-W (7.0%) had more rabbits that were eating than E-F1-S (1.8%; P=0.01). Slaughter lots A-P1-W and C-F1-W had a higher proportion of rabbits that were moving than E-F1-S (both *P*-values<0.08; Table 2). No major difference was observed between slaughter lots for the proportion of rabbits biting or scratching their cage, mating, stretching, shaking, stamping their feet and sneezing. However, E-F1-S was the only one with rabbits that stood up (0.3%).

Of the four rabbit lots from producer D, D-P1-W (51.9%), D-P2-S (60.6%) and D-F2-S (49.0%) had more rabbits that were sitting than D-P1-S (35.7%; all *P*-values<0.04; Figure 1). D-P1-S (62.8%) had more rabbits lying down than D-P1-W (46.9%) and D-P2-S (38.1%; both *P*-values<0.02; Figure 1). With respect to interactions presented in Table 2, D-P1-W (1.9%) had fewer rabbits grooming than D-P1-S (8.0%), D-P2-S (9.5%) and D-F2-S (8.9%; all *P*-values<0.049). No other interactions were different between slaughter lots from producer D.

With respect to season, rabbits were more active in winter than in summer (P=0.045). Animals exhibited fewer interactions in summer (P=0.009). When the animals interacted in winter, none of these interactions were aggressive (P=0.001). Interestingly though, sneezing was more common during the summer (P=0.009).

## Physiological parameters

For rabbits slaughtered in Quebec, we had access to the processing line, which enabled us to measure various physiological parameters. The mean, standard deviation, minimum and maximum of different physiological parameters that were measured are presented in Figure 2. Means for blood lactate ranged from  $0.88\pm0.19$  mmol/L for D-F2-S to  $8.74\pm4.29$  mmol/L for D-P1-S (Figure 2A).

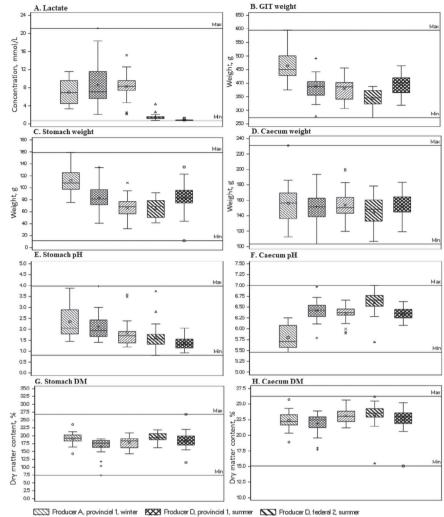
GIT mean weight ranged from  $344.69\pm32.82$  g (D-P2-S) to  $463.85\pm58.53$  g (A-P1-W; Figure 2B). Stomach mean weight ranged from  $65.66\pm18.25$  g (D-P1-W) to  $111.97\pm22.83$  g (AP1-W; Figure 2C). Average caecum weight was

				Producer <sup>2</sup>	3er~					
Variables	A Provincial 1 Winter	B Federal 1 Winter	C Federal 1 Winter	D Provincial 1 Winter	D Provincial 1 Summer	D Provincial 2 Summer	D Federal 2 Summer	E Federal 1 Summer	SEM <sup>3</sup>	<i>P</i> -value
Interaction,%										
Nonaggressive <sup>4</sup>	3.3±0.6 <sup>ab</sup> (1.60-4.9) <sup>5</sup>	3.0±1.0 <sup>ab</sup> (1.2-4.8)	4.0±0.9ª (2.8-5.2)	2.7±0.9 <sup>ab</sup> (1.1-4.2)	1.0±0.3⁰ (0.0-2.5)	2.2±0.6 <sup>ab</sup> (0.0-3.7)	1.3±0.3 <sup>ab</sup> (0.0-2.8)	2.1±0.8 <sup>ab</sup> (0.0-3.8)	0.007	0.05
Aggressive	0.4±0.4 (0.1-0.6)	NO (0.0-0.2)	NO (0.0-0.2)	NO (0.0-0.2)	NO (0.0-0.2)	NO (0.0-0.2)	NO (0.0-0.2)	NO (0.0-0.2)	0.001	Q
No interaction	96.4±0.7 (94.1-98.6)	97.0±1.0 (94.6-99.4)	96.0±0.9 (94.4-97.6)	97.4±0.9 (95.2-99.4)	99.0±0.3 (96.9-100.0)	99.2±1.7 (97.1-100.0)	97.6±1.2 (95.5-99.6)	98.1±0.8 (95.8-100.0)	0.01	Q
Type of activity,%										
Resting	79.8±2.6′ (75.3-84.2)	84.9±1.7 <sup>×y</sup> (80.2-89.6)	84.5±1.5 <sup>w</sup> (81.4-87.6)	89.0±1.6 <sup>×</sup> (84.9-93.1)	83.1±1.6 <sup>w</sup> (79.0-87.2)	81.9±2.2 <sup>w</sup> (77.8-86.0)	82.6±3.0 <sup>w</sup> (78.5-86.6)	87.3±1.9 <sup>w</sup> (82.9-91.8)	0.03	0.07
Drinking	1.6±0.6 <sup>ab</sup> (0.6-2.7)	0.8±0.3 <sup>b</sup> (0.0-1.9)	1.2±0.3 <sup>b</sup> (0.5-1.9)	1.9±0.6 <sup>ab</sup> (0.9-2.9)	1.6±0.3 <sup>ab</sup> (0.6-2.6)	3.8±0.5ª (2.8-4.7)	2.7±0.8 <sup>ab</sup> (1.7-3.7)	0.9±0.3 <sup>b</sup> (0.0-2.0)	0.008	0.001
Grooming	$10.7\pm2.2^{a}$ (7.7-13.7)	4.8±0.9 <sup>ab</sup> (1.6-7.9)	7.0±1.3 <sup>ab</sup> (5.0-9.1)	1.9±0.5 <sup>b</sup> (0.0-4.6)	8.0±1.2ª (5.3-10.7)	9.5±1.3ª (6.8-12.3)	8.9±1.2ª (6.2-11.7)	8.2±1.7ª (5.3-11.2)	0.01	0.001
Biting their cage	NO	NO	0.4±0.3 (0.1-0.7)	NO	NO	0.2±0.2 (0.0-0.6)	0.2±0.2 (0.0-0.6)	0.6±0.3 (0.2-1.0)	0.002	QN
Eating	5.3±1.1 <sup>ab</sup> (3.3-7.3)	7.0±1.1 <sup>a</sup> (4.9-9.1)	4.0±0.9 <sup>ab</sup> (2.6-5.3)	$6.7\pm1.0^{ab}$ (4.8-8.5)	5.2±0.5 <sup>ab</sup> (3.4-7.1)	4.2±0.9 <sup>ab</sup> (2.3-6.0)	3.4±0.9 <sup>ab</sup> (1.6-5.2)	1.8±0.6 <sup>b</sup> (0.0-3.8)	0.01	0.006
Mating	NO	0.2±0.2 (0.0-0.6)	0.6±0.3 (0.3-0.9)	0.1±0.1 (0.0-0.5)	NO	0.3±0.2 (0.0-0.7)	0.1±0.1 (0.0-0.5)	NO	0.002	Q
Moving	2.4±0.7×(1.2-3.4)	2.1±0.7 <sup>w</sup> (0.9-3.2)	2.0±0.6 <sup>x</sup> (1.2-2.7)	0.4±0.3 <sup>w</sup> (0.0-1.4)	1.4±0.4 <sup>w</sup> (0.4-2.4)	0.8±0.3 <sup>%</sup> (0.0-1.8)	0.7±0.3 <sup>w</sup> (0.0-1.7)	NOv	0.006	0.01
Stretching	0.4±0.2 (0.0-0.7)	0.2±0.2 (0.0-0.5)	0.3±0.2 (0.0-0.5)	NO	NO	NO	NO	0.2±0.2 (0.0-0.5)	0.002	Q
Shaking	NO	0.2±0.2 (0.0-0.4)	NO	NO	0.2±0.15 (0.0-0.4)	0.1±0.1 (0.0-0.3)	NO	NO	0.001	Q
Stamping their feet	NO	NO	NO	NO	NO	0.1±0.1 (0.0-0.2)	NO	0.2±0.15 (0.0-0.3)	0.001	Q
Sneezing	NO	NO	NO	NO	0.1±0.1 (0.0-0.3)	0.3±0.2 (0.1-0.6)	0.2±0.2 (0.0-0.4)	0.2±0.2 (0.0-0.4)	0.001	Q
Standing up	NO	NO	NO	NO	NO	NO	NO	0.1±0.2 (0.0-0.3)	0.001	QN
Scratching the cage	NO	NO	NO	NO	0.1±0.1 (0.0-0.2)	NO	NO	0.3±0.2 (0.1-0.5)	0.001	Q

indicates the slaughterhouse inspection jurisdiction at which rabbits were slaughtered. Provincial 1 and 2 were located in Quebec; Federal 1 was located in Untario; Federal 2 is the same slaughterhouse as Provincial 2, but after it received federal accreditation.

<sup>3</sup>SEM: Standard error of the mean.

<sup>4</sup>Offferent letters (a–d) within rows indicate difference at P<0.05 and (x-y) at P<0.10. ND: not different; NO: not observed. Tukey's test was carried out to compare the differences between producers. <sup>5</sup> Confidence interval (lower limit-upper limit).



Producer D, provincial 1, winter
ZZZ Producer D, provincial 2, summer

Figure 2: Physiological parameters (means±standard deviations) measured from rabbits (n = 120) slaughtered in the province of Quebec; blood lactate concentration (A), gastrointestinal tract (GIT) weight (B), stomach weight (C), caecum weight (D), stomach pH (E), caecum pH (F), stomach DM (G) and caecum DM (H). Max = maximum and Min = minimum. Stomach DM and caecum DM = Stomach and caecum dry matter.

relatively stable for all slaughter lots, with means ranging from  $144.92 \pm 19.70$  g for D-P2-S to  $155.90 \pm 26.04$  g for A-P1-W (Figure 2D).

For stomach pH, averages ranged from  $1.37\pm0.29$  for D-F2-S to  $2.35\pm0.73$  for A-P1-W (Figure 2E). Except for A-P1-W, caecum pH was relatively similar for all slaughter lots, with means ranging from  $6.35\pm0.14$  (D-F2-S) to  $6.61\pm0.25$  (D-P2-S; Figure 2F). Slaughter lot A-P1-W was the only one with a mean caecum pH that was below 6 ( $5.80\pm0.26$ ), and the minimum value observed (5.46) was also from this lot. The stomach pH and caecum pH levels from A-P1-W were above and below all of the others, respectively.

Stomach DM ranged from 16.68 $\pm$ 2.81% for D-P1-S to 19.67 $\pm$ 1.72% for D-P2-S (Figure 2G). Caecum DM also was relatively stable for all slaughter lots, with means ranging from 21.88 $\pm$ 1.64% (D-P1-S) to 23.79 $\pm$ 6.35% (A-P1-W; Figure 2H).

FW times observed were categorised into three classes to assess their effect on the parameters measured. Long FW times ( $\geq$ 23 h) were observed for rabbits that were slaughtered outside of Quebec in Flinton Ontario where, unfortunately, we did not have access to the processing line. However, statistical analysis revealed a significant interaction between FW time classes and season for blood lactate concentrations (P=0.005). The concentrations were lowest for rabbits slaughtered in summer after intermediate FW times. Overall, blood lactate concentrations were higher in winter than in summer (P=0.005) and when the FW time was short ( $\leq$ 13.5 h; P=0.002). Stomach pH was also higher when the FW time was short (P=0.047), at 2.23±0.66.

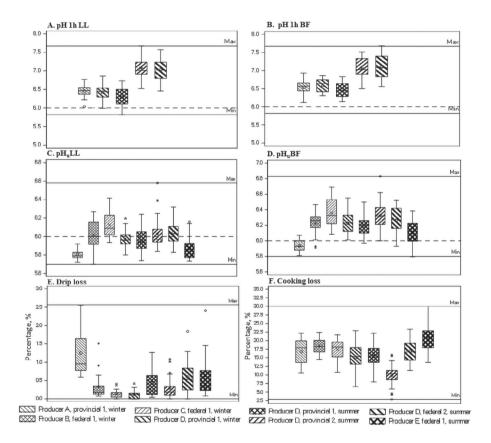


Figure 3: Meat quality characteristics (means±standard deviations) measured from rabbits (n=200) slaughtered in different abattoirs and raised at different farms; pH1hLL (A), pH1hBF (B), pH\_LL (C), pH\_BF (D), drip loss (E) and cooking loss (F). Data for pH1h LL and BF were available only for rabbits slaughtered in Quebec (n=120). Max = maximum and Min=minimum. The pH1hBF and pH\_BF=pH after 1 h and 24 h *post-mortem* of the *Biceps femoris* (BF) muscle, respectively; pH1hLL and pH\_LL=pH after 1 h and 24 h *post-mortem* of the *Longissimus lumborum* (LL) muscle, respectively. Drip loss and cooking loss were from the LL muscle. Dotted lines indicate pH=6.

# Para seguir leyendo, inicie el proceso de compra, click aquí